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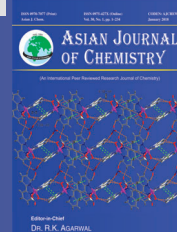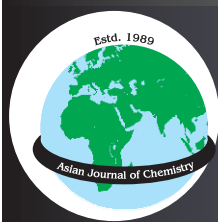Sree Narayana College Chathannur

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Alginate/Chitosan Nanoparticles for Improved Oral Delivery of Rifampicin: Optimization, Characterization and *in vitro* Evaluation

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Alginate/chitosan nanoparticles were synthesized by ionotropic method using natural honey as the stabilizing agent. The nanoparticles were characterized and the potential of these nanoparticles for the controlled oral delivery of antitubercular drug rifampicin was evaluated in terms of entrapment efficiency, swelling behaviour and *in vitro* release of the drug. Carboxyl content and *in vitro* cytotoxicity of the nanoparticles were also evaluated. The swelling and *in vitro* drug release indicates that the system undergoes pH-dependent swelling and release of drug. Drug release was very low in acidic pH and a maximum of only 20 % of the drug was released in 5 h. A sustained release of drug was observed at pH 7.4 and complete release was obtained in 8 h. Kinetics of the drug release was analyzed by fitting the experimental data into Korsmeyer-Peppas equation and show that the mechanism involved in the release was non-Fickian, controlled by a combined mechanism of diffusion and polymer relaxation. This study shows that the obtained nanoparticles can be a potential carrier for the pH controlled oral delivery of rifampicin.

Keywords: Rifampicin, Sodium alginate, Chitosan, Nanoparticle, Honey, Controlled release.

INTRODUCTION

Tuberculosis (TB) caused by *Mycobacterium tuberculosis*, is a chronic communicable and persistent bacterial infection. The current tuberculosis therapy involves the daily adminis-tration of combination of antituberculosis drugs for a period of more than 6 months [1]. But due to limited solubility, rapid and premature degradation and low cellular uptake of these drugs, concentration of the drug reached to target site is less than the therapeutic level. The prolonged drug administration often causes several side effects such as nausea, nephrotoxicity, hepatotoxicity, fever and vomiting. This may results in the patients non-compliance and chance for treatment failure [2]. Treatments based on controlled release drug delivery systems using polymeric micro and nanoparticles is one of the best solution for the problems related to the existing tuberculosis therapy. There are various efforts have been made to develop micro and nano polymer [3]. But some of them include com-plex preparation steps, inconvenient routes of administration and toxicity problems [4]. Oral controlled delivery of rifa-mpicin (RIF) using nano formulation can be improved the efficacy of tuberculosis therapy and the patient comfort. The key factors involved in the selection of carrier in drug delivery

application are its size, shape, drug loading and encapsulation efficiency [5]. Nano particulate drug delivery formulations have the potential to improve these factors and also capable to increase the drug solubility. Alginate a water soluble linear polysaccharide, composed of β-D-mannuronic acid and α-L-guluronic acid (G) is extracted from brown seaweed and is widely used for the bio encapsulation of drugs without the risk of mucosal damage. But, low durability, high-diffusion rates resulting from the high porosity and limited mechanical stabi-lity were the major drawbacks of alginate based drug delivery system [6]. Chitosan, a natural cationic polymer, has the capa-city to interact with anionic bio macromolecules and enhance their loading as holds them tightly. Ribeiro and co-workers could successfully retarded the release of haemoglobin from alginate micro particles by chitosan coating [7]. The studies made by Wittaya-areekul *et al.* [8] and Takka & Gürel [9] revealed that the incorporation of chitosan into alginate micro particles enhances its bioadhessive nature. Oral delivery with alginate nanoparticles coated with chitosan is expected to offer advantages of controlled drug delivery, high drug payload and improved stability. It also ensure protection for encapsulated drugs from acidic environment of gastrointestinal tract and enzyme degradation. The small size of nanoparticles increases

the specific surface area and allowing increased contact area with the epithelial surface and results in greater uptake of drugs. However, the formulation and application of alginate-chitosan nanoparticles, have not gained much attention as the alginate-chitosan microparticles. Honey has antibacterial and antifungal properties and has been used in Ayurveda medicine of India, as a vehicle for faster absorption of various drugs such as herbal extracts. It also supports the treatment of several specific ailments, particularly those related to respiratory irritations and infections, mouth sores and eye cataracts [10,11]. The objective of the present work was to develop alginate-chitosan (ALG-CH nanoparticles) using natural honey as stabilizer, an ideal drug delivery system that is potentially expected to provide pH dependent drug release, with increased bioavailability of the drug, prolonged circulation time and reduced side effects. The nanoparticles developed were loaded with rifampicin and characterized for the application towards its utilization as a carrier for rifampicin. *In vitro* release kinetics from these nanoparticles was studied. From the present study, it is found that ALG-CH nanoparticles are excellent carrier system for the controlled delivery of rifampicin.

EXPERIMENTAL

Low viscosity sodium alginate powder (Sigma-Aldrich, London), chitosan (Sigma-Aldrich, London), rifampicin (Himedia Laboratories, Nasik), calcium chloride dihydrate (Merck, Germany) were of analytical grade and used as such without any further purification. Natural honey used in this study was procured from Kerala Agriculture University.

Preparation of alginate nanoparticle: Alginate-chitosan nanoparticles were prepared by ionotropic gelation technique with slight modification. Sodium alginate was dissolved in aqueous honey (added as surfactant and stabilizer) solution (1 %, w/v). Chitosan was dissolved in aqueous acetic acid (0.25 %, w/v) and is added to sodium alginate solution. It was followed by the addition of rifampicin (0.5 %,w/v). Aqueous calcium chloride solution (1 %, w/v) was added drop-wise into this solution with continuous stirring using a magnetic stirrer. pH of the entire solution was maintained at 5.5. This homogenized mixture was sonicated for 5 min. Nanoparticles were collected by centrifugation at 3500 rpm for 5 min, washed and dried under vacuum [12]. The ratio of alginate to chitosan was optimized on the basis of their effect on drug entrapment efficiency.

Material characterization methods: Fourier transform infrared (FTIR) spectra of alginate, chitosan, rifampicin, ALG-CH nanoparticles and rifampicin loaded ALG-CH nanoparticles were recorded between 4000 and 400 cm^{-1} wavelength range using Shimadzu FTIR model 1801. Surface morphology of the ALG-CH nanoparticles was probed using a JEOL JSM-6390LA analytical scanning electron microscope. Transmission electron microscopy (TEM) was performed using JEOL model 1200EX instrument operated at an accelerating voltage at 80 kV. XRD patterns of the samples were recorded by using Brucker D8 Advance diffractometer with monochromatic Cu-Kα_1 radiation (λ = 1.5418 Å).

Swelling behaviour: The swelling behaviour of nanoparticles were determined by tea-bag method [13]. A known amount of dried sample (W$_d$) was taken in a previously weighed

tea bag (of suitable mesh size) and allowed to swell in solution of pH 1.2 and 7.4 respectively for a specific period. The swollen samples were taken out and weighed immediately after removing the excess liquid from the surface using filter paper (W$_s$). The percentage of swelling was calculated using the equation. Data were given as mean ± standard deviation (SD) based on 3 independent measurements.

$$SD = \left(\frac{W_s - W_d}{W_d} \right) \times 100 \qquad (1)$$

Estimation of carboxyl group: Carboxyl groups present in ALG-CH nanoparticles before and after cross linking was determined by the Boehm titration method [14]. About 0.1 g sample was shaken with sodium bicarbonate (30 mL, 0.1 M) for 20 h at room temperature and filtered. Excess HCl (0.05 M) was added to 10 mL of filtrate, boiled off CO$_2$, cool the solution and back titrated with NaOH (0.05 M). The amount of carboxyl group was calculated using the equation.

$$COOH\left(\frac{meq}{g} \right) = \frac{(C_{NaOH} \times V_{NaOH} - C_{HCl} \times V_{HCl})}{W} \qquad (2)$$

Cytotoxicity evaluation: The *in vitro* toxicity of ALG-CH nanoparticles were measured using L929 fibroblast by MTT assay. The cells were cultured with DMEM containing 10 % FBS and the cultured cell lines were kept at 37 °C in a humidified 5 % CO$_2$ incubator for 24 h. The medium was then replaced with test solution of different concentration (100 µg, 50 µg, 25 µg, 12.5 µg, 6.25 µg/mL) and incubated at 37 °C in a humidified 5 % CO$_2$ incubator for specified time. Later 30 µL of MTT (5 mg/mL) was added to the wells and incubated at 37 °C in a humidified 5 % CO$_2$ incubator for 4 h. After the incubation period, the supernatant was removed and 100 µL DMSO was added to solubilize the formazan crystals. The absorbance values were measured by using micro plate reader at a wavelength of 570 nm to determine the optical density (OD). Optical density of control was obtained from the cells by repeating the above experiment without any sample. Cell viability (%) was calculated using eqn. 3.

$$Cell\ viability\ (\%) = \left(\frac{OD\ of\ sample}{OD\ of\ control} \right) \times 100 \qquad (3)$$

Determination of entrapment efficiency of nanoparticles: The concentration of non-encapsulated rifampicin was determined by the absorbance at 475 nm, using a UV-visible spectrophotometer (Perkin Elmer Lambda Bio 40). The drug entrapment efficiency (% EE) was calculated by eqn. 4 as follows:

$$EE\ (\%) = \left(\frac{Weight\ of\ RIF\ in\ ALG\text{-}CH\ NPs}{Weight\ of\ RIF\ used\ for\ loading} \right) \times 100 \qquad (4)$$

Data were given as mean ± standard deviation (SD) based on 3 independent measurements.

***In vitro* drug release:** *In vitro* drug release studies were carried out in simulated gastric fluid (SGF, pH 1.2) and simulated intestinal fluid (SIF, pH 7.4) prepared according to the United States Pharmacopoeia at 37 °C under shaking conditions [15]. Aliquots were drawn at specific time intervals and replaced with equal volume of buffer. The rifampicin-loaded nanoparticles were separated from the aqueous suspension by

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ultra-centrifugation at 10,000 rpm for 10 min (SORALL® Bioguge Stratos Ultracentrifuge). Rifampicin's concentration was determined by the absorbance at 475 nm in a UV-visible spectrophotometer (Perkin Elmer Lambda Bio 40). Data were given as mean ± standard deviation (SD) based on 3 independent measurements.

Drug release kinetics: To find out the drug release mechanism, the drug release data was analyzed using Korsmeyer-Peppas equation eqn. 5.

$$\frac{Q_t}{Q_\infty} = kt^n \tag{5}$$

Q_t/Q_∞ is fraction of drug released at time t, K is the rate constant and n is the release exponent. In this model, the value of n identifies the release mechanism of drug. The values of these parameters vary with the geometric form of the system, that is, thin film, cylindrical or spherical. For a spherical system ≤ 0.43 indicates Fickian release and n = 0.85 indicates a purely relaxation-controlled delivery which is referred to as Case II transport. Intermediate values $0.43 < n < 0.85$ indicate an anomalous behaviour (non-Fickian kinetics) corresponding to coupled diffusion/polymer relaxation been observed [16-18].

RESULTS AND DISCUSSION

Preparation and characterization of rifampicin loaded alginate nanoparticle: Hybrid nanostructures of natural polymers are of broad technological interest in drug delivery applications as they are nontoxic and biocompatible in nature. Hybrid structures offer the possibility of tuning release characteristics and pharmacological performance by varying the concentration and functional properties of the components. Here a green method was adopted for the preparation of ALG-CH nanoparticles using water as solvent and honey as the stabilizing agent. Formation of nanoparticle was found to be dependent on the concentration and pH of the solution. Critical condition for nano formation was found to be at 4:1 alginate-chitosan ratio and a pH of 5.5. The results of optimized parameters are summarized in Table-1.

TABLE-1
ENTRAPMENT EFFICIENCY OF VARIOUS
FORMULATIONS OF ALG-CH NANOPARTICLE

ALG:CH (weight ratio)	Entrapment efficiency (%)
1:1	42.31 ± 1.9
2:1	45.76 ± 3.5
3:1	50.87 ± 2.8
4:1	55.62 ± 2.6
5:1	48.56 ± 3.1

On the addition of calcium chloride, the Ca^{2+} induced interchain association *via* carboxyl group of G units, leading to the formation of junction zones [19,20]. Unlike other organic surfactants, here natural honey was used as the stabilizing agent [21]. FTIR was used to characterize the potential interactions in the nanoparticles. The FTIR spectrum of ALG, CH, rifampicin, neat ALG-CH nanoparticle and rifampicin loaded ALG-CH nanoparticle are given in Fig. 1. The carboxylate groups of alginate associated with the amino groups of chitosan through electrostatic interactions to form the polyelectrolyte
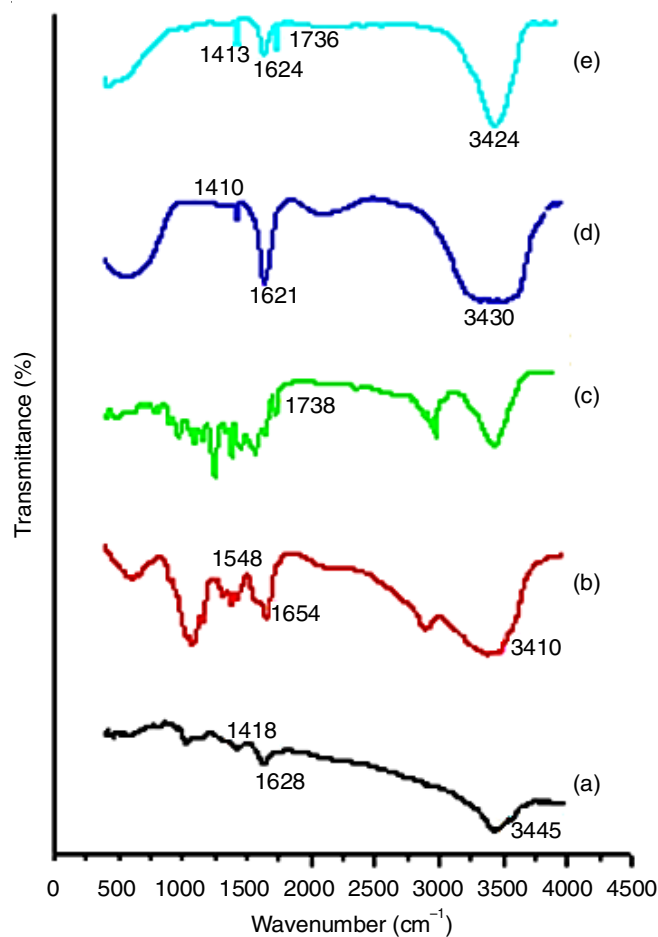

Fig. 1. FTIR of (a) alginate (b) chitosan (c) rifampicin (d) ALG-CH (e) rifampicin loaded ALG-CH nanoparticle

complex. This is confirmed by the notable shift in the position of the amide I vibration (1654 to 1621 cm⁻¹) of chitosan. The characteristic peak of rifampicin at 1741 cm⁻¹ corresponds to furanone ring structure. This band can also be seen in the rifampicin loaded ALG-CH nanoparticle, indicating that the rifampicin molecule was incorporated into the polymeric network of the nanoparticle and there is no interaction between the drug and polymeric network [22,23].

The shape and surface morphology of the system was characterized by SEM and TEM and is given in Fig. 2. The SEM images show the presence of well-defined spherical nanoparticles. It was confirmed from TEM and indicates the presence of nanoparticles of size less than 100 nm. The presence of honey help to reduce the particle size. The XRD patterns of the rifampicin and rifampicin loaded ALG-CH nanoparticles is shown in Fig. 3. The powder X-ray diffraction of rifampicin revealed its crystalline character showing major peaks at 2θ = 10.7, 15.4, 16.6, 17.6, 19.5, 21, 22.7, 25, 27 and 29. In rifampicin loaded ALG-CH nanoparticle, the disappearance of the crystalline peaks of the drug is indicative of its actual amorphization during the nanoparticle preparation.

Swelling behaviour: The swelling behaviour of ALG-CH nanoparticles were determined in simulated gastric fluid (SGF, pH 1.2, Fig. 4 a) and intestinal fluid (SIF, pH 7.4, Fig. 4b) at room temperature. Swelling behaviour of nanoparticle has a profound effect on the mechanism and kinetics of drug release.
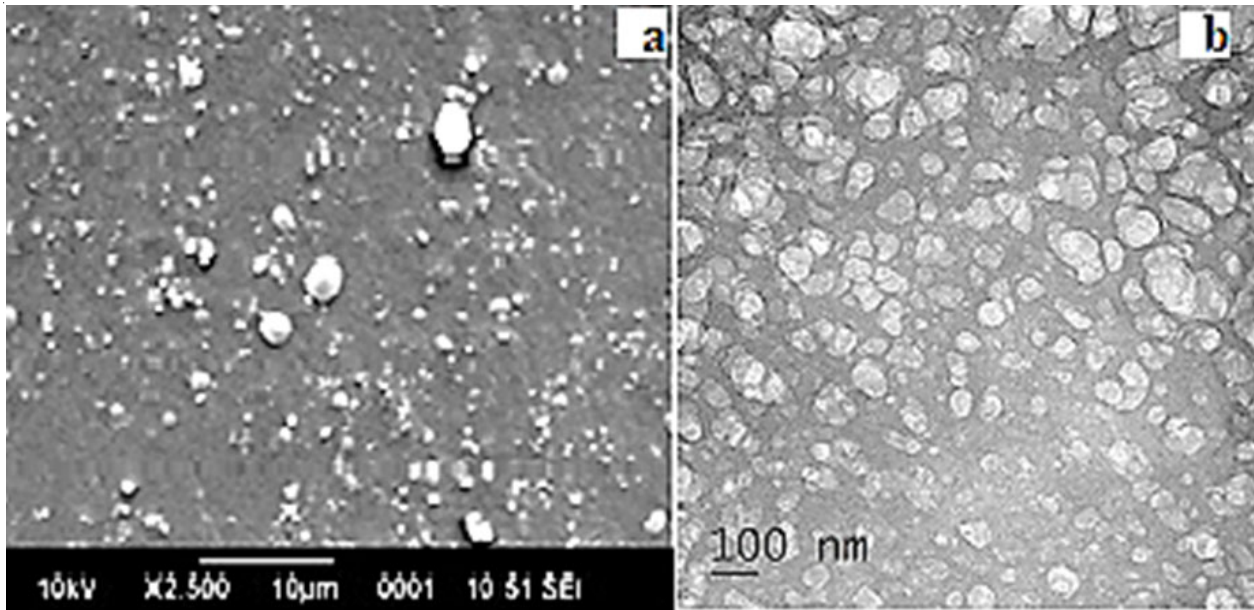
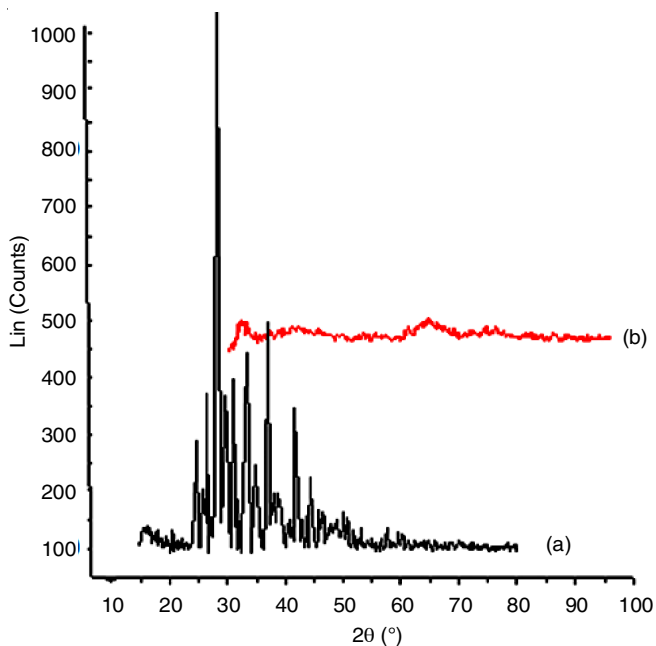Fig. 2. (a) SEM and (b) TEM images of ALG-CH nanoparticle

Fig. 3.　XRD pattern of (a) rifampicin and (b) rifampicin loaded ALG-CH nanoparticle
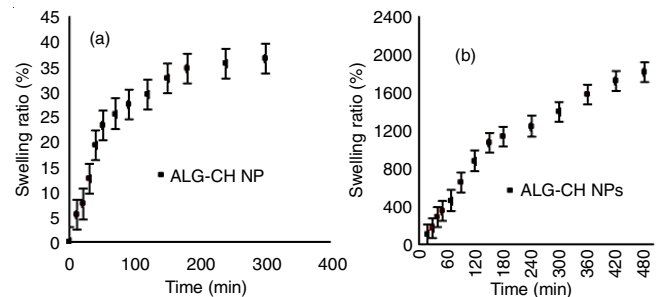

Fig. 4.　Swelling behaviour of ALG-CH nanoparticles in (a) pH 1.2 (b) pH 7.4

in nanoparticles before and after cross linking were found to be 3.15 meq g^{-1} and 2.20 meq g^{-1} respectively.

Cytotoxicity: The cytotoxicity of ALG-CH nanoparticle was evaluated by MTT assay on L929 cell lines at different concentrations and the results shown in Fig. 5. From this graphical representation, it was found that the obtained nanoparticle has no toxicity and suited for drug delivery applications.

***In vitro* drug release:** The *in vitro* drug release studies were performed in the fluids simulating the conditions of

At low pH (pH 1.2) the swelling was very low. In acidic medium, the carboxylate groups of alginate are protonated and are converted to insoluble alginic acid. The electrostatic repulsion among carboxylate groups reduces swelling. The disappearance of electrostatic repulsions favours the shrinkage of nanoparticle. The percentage swelling of nanoparticles were higher at pH 7.4. In ALG-CH nanoparticles there exists some electrostatic interactions between the –NH$_3^+$ groups of chitosan chains and the –COO$^-$ groups of alginate. It provides a strong and compact layer and improves the stability of ALG-CH nanoparticles and ultimately prolonging the time for degradation and collapse. These observations showed that ALG-CH nanoparticles undergoes pH-dependent swelling, in which the carboxylate and amino groups respond to stimuli from the environment in which they are inserted. The amount of the free carboxyl group
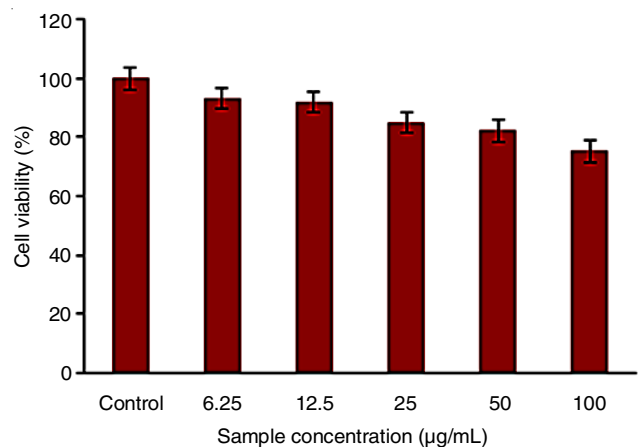

Fig. 5. *In vitro* cell viability of L929 cells treated with ALG-CH nanoparticle

gastrointestinal (GI) tract for 0-5 h in SGF (pH 1.2) and 0-8 h in SIF (pH 7.4) Fig. 6. The amount of rifampicin released in SGF at pH 1.2 is very low. Only 15-20 % of rifampicin is released from ALG-CH nanoparticle. At this pH, these nanoparticles were not eroded, the negative –COO group of calcium alginate protonated and acted as a diffusion barrier for the release of the drug. *In vitro* drug release was higher in the SIF compared to the SGF. At pH 7.4 a rapid increase in the release rate was observed up to 100 % in 8 h. This can be attributed to the introduction of chitosan that may have restricted the motion of the alginate polymer chains and thereby improving the entrapment and controlled release behaviours. Chitosan also acts as a reinforcing polymer to retard the erosion of alginate nanoparticles, which explains the slow, but sustained *in vitro* drug release.
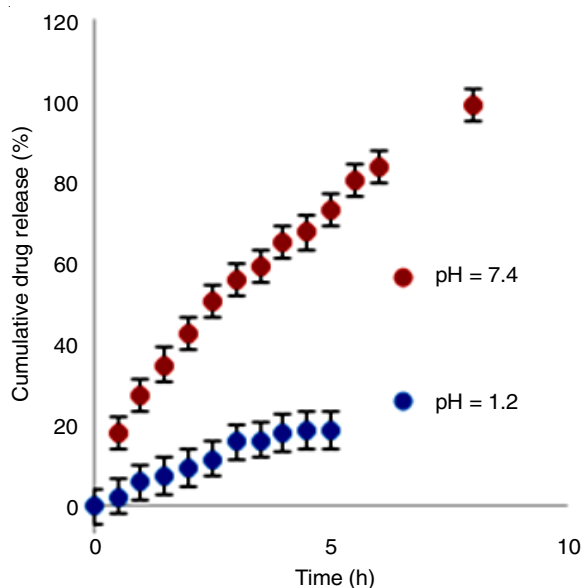

Fig. 6.   Rifampicin cumulative release from ALG-CH nanoparticle in pH 1.2 and 7.4

Drug release kinetics: The drug release kinetics of rifampicin from nanoparticles was analyzed by fitting the release data with Korsmeyer-Peppas model. The value of the release exponent n was 0.77 for ALG-CH nanoparticles which indicate an anomalous, non-Fickian kinetics corresponding to coupled diffusion/polymer relaxation mechanism.

Conclusion

Alginate-chitosan nanoparticles containing rifampicin were obtained successfully by simple ionotropic gelation method using natural honey as stabilising agent. The morphological analysis carried out by SEM and TEM revealed that the polymers used in the formulations conferred particular surface characteristics to the polymeric nanoparticles. The drug release and swelling were pH-dependent. The drug was well protected in the acidic environment of gastric fluid and released completely in intestinal fluid. *In vitro* cytotoxicity studies indicate that the nanoparticles are biocompatible. The values obtained for the release parameters indicate an anomalous transport mechanism. This study demonstrates that the nanoparticles developed provide a good system for the controlled-release of rifampicin.

REFERENCES

1. E. Nuermberger and J. Grosset, *Eur. J. Clin. Microbiol. Infect. Dis.*, **23**, 243 (2004);
    https://doi.org/10.1007/s10096-004-1109-5.
2. R. Parmar, R. Misra and S. Mohanty, *Colloids Surf. B: Biointerfaces*, **129**, 198 (2015);
    https://doi.org/10.1016/j.colsurfb.2015.03.051.
3. D. He, P. Deng, L. Yang, Q. Tan, J. Liu, M. Yang and J. Zhang, *Colloids Surf. B: Biointerfaces*, **103**, 580 (2013);
    https://doi.org/10.1016/j.colsurfb.2012.10.062.
4. P. O'Hara and A.J. Hickey, *Pharm. Res.*, **17**, 955 (2000);
    https://doi.org/10.1023/A:1007527204887.
5. W. Liu, W.D. Wu, C. Selomulya and X.D. Chen, *Powder Technol.*, **236**, 188 (2013);
    https://doi.org/10.1016/j.powtec.2012.02.012.
6. K. Moebus, J. Siepmann and R. Bodmeier, *Eur. J. Pharm. Biopharm.*, **72**, 42 (2009);
    https://doi.org/10.1016/j.ejpb.2008.12.004.
7. A.J. Ribeiro, C. Silva, D. Ferreira and F. Veiga, *Eur. J. Pharm. Sci.*, **25**, 31 (2005);
    https://doi.org/10.1016/j.ejps.2005.01.016.
8. S. Wittaya-areekul, J. Kruenate and C. Prahsarn, *Int. J. Pharm.*, **312**, 113 (2006);
    https://doi.org/10.1016/j.ijpharm.2006.01.003.
9. S. Takka and A. Gürel, *AAPS PharmSciTech*, **11**, 460 (2010);
    https://doi.org/10.1208/s12249-010-9406-z.
10. S.K. Motwani, S. Chopra, S. Talegaonkar, K. Kohli, F.J. Ahmad and R.K. Khar, *Eur. J. Pharm. Biopharm.*, **68**, 513 (2008);
    https://doi.org/10.1016/j.ejpb.2007.09.009.
11. R. Singh and J.W. Lillard Jr., *Exp. Mol. Pathol.*, **86**, 215 (2009);
    https://doi.org/10.1016/j.yexmp.2008.12.004.
12. P. Geetha, M.S. Latha, S.S. Pillai and M. Koshy, *Ecotoxicol. Environ. Saf.*, **122**, 17 (2015);
    https://doi.org/10.1016/j.ecoenv.2015.06.032.
13. E. Al, G. Güçlü, T.B. Iyim, S. Emik and S. Özgümüs, *J. Appl. Polym. Sci.*, **109**, 16 (2008);
    https://doi.org/10.1002/app.27968.
14. T.S. Anirudhan, Binusreejayan and J.R. Deepa, *J. Appl. Polym. Sci.*, **133**, 434 (2016);
    https://doi.org/10.1002/app.43479.
15. M.D. Rockville, The United States Pharmacopeial Convention, US Pharmacopeia, **XXVI/NF**, 2528 (2003).
16. R.W. Korsmeyer, R. Gurny, E. Doelker, P. Buri and N.A. Peppas, *Int. J. Pharm.*, **15**, 25 (1983);
    https://doi.org/10.1016/0378-5173(83)90064-9.
17. P.L. Ritger and N.A. Peppas, *J. Control. Rel.*, **5**, 23 (1987);
    https://doi.org/10.1016/0168-3659(87)90034-4.
18. P.L. Ritger and N.A. Peppas, *J. Control. Rel.*, **5**, 37 (1987);
    https://doi.org/10.1016/0168-3659(87)90035-6.
19. P. Sikorski, G.Skjåk-Bræk and B.T. Stokke, *Biomacromolecules*, **8**, 2098 (2007);
    https://doi.org/10.1021/bm0701503.
20. E.V.R. Campos, N.F.S. de Melo, E. de Paula, A.H. Rosa and L.F. Fraceto, *J. Colloid Sci. Biotechnol.*, **2**, 106 (2013);
    https://doi.org/10.1166/jcsb.2013.1040.
21. J.S. Yang, H.B. Ren and Y.J. Xie, *Biomacromolecules*, **12**, 2982 (2011);
    https://doi.org/10.1021/bm200571k.
22. C. Sartori, D.S. Finch, B. Ralph and K. Gilding, *Polymer*, **38**, 43 (1997);
    https://doi.org/10.1016/S0032-3861(96)00458-2.
23. L. Lacerda, A.L. Parize, V. Fávere, M.C.M. Laranjeira and H.K. Stulzer, *Mater. Sci. Eng.*, **39**, 161 (2014);
    https://doi.org/10.1016/j.msec.2014.01.054.

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