



## Identification of Antimastitis Componentes in *Boerhavia diffusa* as an Inhibitor of *Staphalococcus aureus* Monofunctional Glycosyltransferase, Causing *Bovine mastitis* (An Insilico Approach)

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<http://dx.doi.org/10.13005/ojc/340264>


(Received: December 08, 2017; Accepted: January 20, 2018)

### ABSTRACT

*Bovine mastitis* is an infection of cattle leading to a huge reduction in milk production that causes severe economic loss in dairy industry across the world. Causative of the disease includes bacteria, virus and non-bacterial pathogens. Among these, *Staphalococcus aureus* is the common cause of mastitis and is highly resistant to the most routinely used antibiotics. So current antibiotic therapy has shown limited efficacy. The crude extract of locally available medicinal plant *Boerhavia diffusa* is used traditionally against mastitis and is found to be highly effective. The objective of the study is the identification of the phytochemicals in *Boeravia diffusa* responsible for the antimastitis activity by insilico docking analysis using Schrodinger Suit v 9.2. 20 phytochemicals in *Boerhavia diffusa* were selected for docking studies based on ADMET properties. The high resolution crystal structure of the target receptor protein of *Staphalococcus aureus* was retrieved from PDB. Structure of the phytochemicals and the most commonly used antibiotic against *Bovine mastitis* were selected from PUB CHEM NCBI. The phytochemicals, Boeravinone A, B, C, D, E and F from *Boehravia diffusa* showed good docking scores and better interaction with the active sites of the target proteins used for the evaluation than the most commonly used commercially available drug. Results of this study are important for the designing of novel drugs for the treatment of mastitis.

**Keywords:** *Staphalococcus aureus* monofunctional glycosyltransferase, *Bovine mastitis*, Docking, *Boerhavia diffusa*, Qik prop.



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## INTRODUCTION

*Bovine mastitis* is a kind of inflammation of the udder of cattle due to wounds or scars. It is associated with the presence of infectious agents as bacteria, Bovine Herpes virus, non bacterial pathogens like mycoplasmas, fungi, yeast and chlamydia<sup>1-2</sup>. Among these, bacterial infections are most common. *Bovine mastitis* reduces the quantity of casein, lactoferrin and potassium in milk. As the major protein, casein in milk deteriorates, the calcium level in milk also decreases. During processing and storage also the milk protein undergo deterioration<sup>3</sup>. Milk from affected animals shows very high somatic cell count which lowers the quality of milk<sup>4</sup>. Hence, *Bovine mastitis* causes severe economic losses in dairy industry.<sup>5</sup>

Current methods used in the control of *Bovine mastitis* are based on antibiotic therapy. The commonly used antibiotics are penicillins and semisynthetic penicillin derivatives targeting the penicillin binding proteins of *Staphalococcus aureus*. Penicillin binding proteins or DD- transpeptidase form bonds between the oligopeptide crosslinks in peptidoglycan<sup>6</sup>. Inhibition of peptidoglycan synthesis leads to bacterial cell lysis<sup>7</sup>. Commonly used antibiotics target the DD-transpeptidase and inhibit peptidoglycan synthesis there by leading to bacterial cell lysis. But mutations in genes coding for transpeptidases leads to reduced interaction with antibiotic results in the generation of antibiotic resistance in *Staphalococcus aureus*<sup>8</sup>. This is the main reason for the decreased antibiotic efficacy in the treatment of *Bovine mastitis*. Monofunctional glycosyl transferase<sup>1</sup> is an alternate potent target of *Staphalococcus aureus*, the inhibition of which also leads to bacterial cell lysis.

The immunomodulator compounds derived from medicinal plants has potent application in controlling mastitis. *Boerhavia diffusa* is one of the medicinal plants in the whole or its peculiar parts have enormous medicinal properties<sup>9</sup>. Farmers of the Southern region of Kerala uses the crude extract of the plant as a whole as a medicine for mastitis from their traditional knowledge.

## Methodology

### Bioinformatics analysis

#### Preparation of protein

Crystal structure of the target proteins of *Staphalococcus aureus* Monofunctional Glycosyltransferase was obtained from RCSB Protein Data Bank (PDB ID: 3HZS)<sup>10</sup>. Automatically imported PDB files from the RCSB PDB website to the Maestro working interface.<sup>10-12</sup> Optimized the protein's hydrogen bond network by means of a systematic, cluster-based approach, which greatly decreased preparation times and then performed a restrained minimization that allows hydrogen atoms to be freely minimized. Here the crystal structure of the target protein was complexed with a ligand. So sitemap creation was exempted.

#### Ligand preparation

Twenty phytochemicals present in *Boerhavia diffusa*<sup>13</sup> were selected to find out the inhibitory activity towards the target protein. (Betasitosterol, BoeravinoneA, B, C, D, E, F, Hexacosanoic acid, Tetracosanoic acid, Sitosterylferulate, Liriodendrin, Arachidic acid, Triaccondanol, Palmitic acid, Stearic acid, Sitosterylpalmitate, Hentriacontane, Sitosterylester, Ursolic acid). The structure of the phytochemicals and commonly used commercially available antibiotic Cloxacillin were downloaded from pub chem in the (.sdf) format. These ligands were subjected to ligand preparation using the ligand preparation wizard (ligprep) of Schrodinger software in the *Maestro interface*<sup>12</sup><sup>11-12</sup>. One low energy conformation was generated. The ligprep ligands were used for the Docking analysis.

#### Docking studies

The compounds were screened by Schrodinger docking software to study inhibitors of the target protein. Grid generation was done using the centroid of workspace ligand R0 48-8071. The rigid receptor docking using the Glide program was carried out against the target protein with the set of ligands. The mode of docking was selected as XP (Extra precision) for a high docking accuracy. The glide docking was carried out for the minimised protein<sup>11,12,14</sup>.

### Ligand interaction study

The ligand molecules fit in to the active binding sites of the protein molecules by means of some interactions. The interactions include hydrogen bonding, pi-pi stacking interactions and pi-pi cation interactions. A detailed information of various interactions of the ligands with amino acid residues pointing the type of bonds, bond length and various angles were studied using this option.

### ADMET properties prediction

The bioactive compounds from *Boerhavia diffusa* were checked for their ADMET properties using Qik prop module.<sup>14-15</sup> QikProp helped in analyzing the pharmacokinetics and pharmacodynamics of the ligand by accessing the drug likeness.

## RESULTS AND DISCUSSION

*Boerhavia diffusa* is a renowned medicinal plant due to its therapeutic potential. Aqueous leaf extract of *Boerhavia diffusa* had hypoglycaemic effect<sup>16</sup> and nutritive properties<sup>17</sup>. Ethanolic extract of *Boerhavia diffusa* leaves had antistress, adoptogenic, immunopotentiating<sup>18</sup> and anti-inflammatory activity<sup>19</sup>. Antibacterial activity of methanolic and ethanolic extracts of *Boerhavia diffusa* leaves against both gram positive and Gram-negative bacteria were reported.<sup>20</sup> Also aqueous and ethanolic extracts of *Boerhavia diffusa* had antibacterial activity on *E.Coli*, *Staphalococcus aureus* and *P. aeruginosa*.<sup>21</sup>

Present study revealed the phytochemicals in *Boerhavia diffusa* which are responsible for the antimastitis activity by inhibiting the most potent drug target in *Staphalococcus aureus*. Table 1 shows the docking scores, number of hydrogen bonds, interacting amino acid residues in the active site of protein and hydrogen bond length. Table 2 explains the drug likeness of phytochemicals predicted by QikProp simulation. Fig. 1 and Fig. 2 shows the 2D and 3D interactions of the ligands with the target protein.

From the results obtained, it has been understood that there exist excellent binding interactions between phytochemicals in *Boerhavia diffusa* and target proteins (PDB ID:4HZS)

compared to the commercial drug (cloxacillin) with good binding scores. In the protein monofunctional glycosyltransferase with PDB ID: 3HZS, the 110<sup>th</sup>PHE, 136<sup>th</sup>GLN and 130<sup>th</sup>GLN amino acid residues forms the binding pocket. The phytochemical Boeravinone E shows a binding score -6.151 kCal/mol and forms three hydrogen bonds with the amino acid residues in the binding pocket. The hydroxyl group in the 11<sup>th</sup> position of the chromeno chromon nucleus forms a hydrogen bond with the amino acid residue 110<sup>th</sup> PHE (bond length-2.04799, Acceptor Angle-143.297 and Donor angle-138.404) Similarly, the hydroxyl group in the 9<sup>th</sup> position of the chromeno chromon nucleus of Boeravinone E forms a hydrogen bond with 130<sup>th</sup>GLY residue. (Bond length-2.18231, Acceptor angle-143.17 and Donor angle-119.565) There is a side to side hydrogen bond between the ketonic oxygen of Boeravinone E with 136<sup>th</sup>GLN amino acid residue. (bond length-2.15438, Donor angle-162.547).

The phytochemical Boeravinone D exhibit three hydrogen bonding interactions with a docking score -6.013kCal/mol. The two hydroxyl groups in the 11<sup>th</sup> and 9<sup>th</sup> positions of the chromeno chromon nucleus forms two hydrogen bonds with the 130<sup>th</sup> GLY and 110<sup>th</sup>PHE amino acid residues. (Bond length-2.11767 and 1.89519, Acceptor angle-170.817 and 151.217, Donor angle-166.252 and 138.416) respectively. The ketonic oxygen of the chromon nucleus forms a side to side hydrogen bond with 136<sup>th</sup>GLN. (Bond length-2.09866, Donor angle-165.062).

Boeravinone F forms four hydrogen bonds with a docking score -5.629. The hydroxyl group in the 11<sup>th</sup> position forms a hydrogen bond with the amino acid residue 136<sup>th</sup>GLN (Bond length-2.14702, acceptor angle-170.600 and Donor angle-149.579). This hydroxyl group also forms another hydrogen bond with 110<sup>th</sup>PHE (Bond length-1.88670, acceptor angle-161.487 and Donor angle-156.617. Ketonic oxygen forms a side to side hydrogen bond with 136<sup>th</sup> GLN amino acid residue.

Boeravinone A forms two hydrogen bonds with docking score -5.575kCal/mol. The hydroxyl

group in the 9<sup>th</sup> position of the chromeno chromon nucleus of Boeravinone A forms a hydrogen bond with 130<sup>th</sup>GLN residue (bond length-2.23410, Acceptor angle-173.152 and donor angle-164.273) Hydroxyl group in the 11<sup>th</sup> position forms one side

to side hydrogen bond with 110<sup>th</sup>PHE. (bond length-1.97871, Acceptor angle-165.258, donor angle-173.134) The ketonic oxygen forms a hydrogen bond with 136<sup>th</sup>aminoacid residue. (bondlength-2.17994, Acceptor angle-158.265 and donor angle-178.365)

Boeravinone C shows a docking score -4.528kCal/mol and forms one hydrogen bond between the hydroxyl group and 130<sup>th</sup>GLY aminoacid residue. (bond length-1.83269, acceptor angle-165.235 and donorangle-149.245). All these components satisfied the ADME properties (Table 2).

**Table 1: The phytochemicals in *Boerhavia diffusa* having good docking result with *Staphalococcus aureus* mon of uncton alglycosyl transferase**

Ligands	Pub Chem ID of ligands	Docking Score (Kcal/mol)	Number of hydrogen bond	Interacting aminoacid residues of active site	Hydrogen bond length(A <sup>o</sup> )
Protein (PDB ID : 3HZS)					
Boeravinone E	11537197	-6.151	3	110 PHE,136GLN,130 GLY 130 GLY	2.04799,2.15438 2.18231
Boeravinone D	15081178	-6.013	3	110 PHE,136GLN, 130 GLY	1.89519,2.09866 2.11767
Boeravinone F	12004175	-5.629	4	110 PHE,136GLN, 136 GLN,130GLY	1.88670,2.147022,77985, 2.30043
Boeravinone A	14018346	-5.575	3	110PHE,136GLN, 130GLY	1.97871,2.17994 2.23410
20-hydroxy ecdysone	5459840	-5.163	4	131GLY,136GLN	1.88383,2.180481,97645, 1.7501009
Boeravinone C	13940643	-4.528	1	110PHE,130GLY 130GLY	1.7501009 1.83269
Cloxacillin	6098	-4.256	2	123GLN,136GLY	2.06780,2.57157

**Table 2: Prediction of drug- likeness of phytochemicals using qik prop simulation<sup>14</sup>**

Ligand(Phytochemicals from <i>Boerhavia diffusa</i> )	Molecular Weight (MW)g/mol	Hydrogen bond donor	Hydrogen bond acceptor	Q <sub>p</sub> LogP <sub>(o/w)</sub>
Boeravione C	344.319	2.000	5.500	2.215
Boeravione D	342.303	2.000	6.200	1.734
Boeravione B	312.277	1.000	6.250	1.701
Boeravione A	326.304	1.000	5.450	2.565
Boeravione F	326.260	2.000	6.250	0.920
Boeravione E	328.276	3.000	6.200	0.967
Cloxacillin	435.879	5.000	11.300	0.283

(Range of properties - Molecular weight less than 500, Hydrogen bond donor 0-6, Hydrogen bond acceptor 2-20 and Q<sub>p</sub>logP<sub>(o/w)</sub> -2 - 6.5)





inhibitory activity towards the *staphalococcus aureus* monofunctional glycosyltransferase compared to the commercial drug Cloxacillin. These phytochemicals are responsible for the antimastitis activity of *Boerhavia diffusa*. This study leads to the development of novel drugs for mastitis.

## REFERENCES

1. Reena Patel, J.; Prashant Kunjadia, D.; ChaitanyaJoshi, G.; Anju Kunjadia, P. *International Journal of Pharmacy and Pharmaceutical sciences.*, **2015**, *7*, 496-502.
2. Wellenberg, G.J.; Van der Poel, W.H.M.; Van Orischo, J.T. *Veterinary Microbiology.*, **2002**, *88*, 27- 45.
3. Harmon, R.J. *J. Dairy Sci.*, **1994**, *77*, 2103-2112.
4. Kandasamy, S.; Green, B.B.; Benjamin, A.L.; Kerr, D.E. *J Dairy Sci.*, **2011**, *12*, 5963-75.
5. *Department of Animal Science*. "Mastitis in Dairy Cows" (PDF). *MacDonald Campus of McGill University. Archived from the original (PDF) on July 8, 2003*. Retrieved 4 February **2010**.
6. <http://en.wikipedia.org/wiki/peptidoglycan>
7. Andrea FeBler;Carly Scott; Kristina Kadlec; Ralf Ehricht; Stefan Monecke; Stefan Schwarz. *Journal of Antimicrobia chemotherapy.*, **2010**, *65*, 619-625.
8. Muhamed Mubarack, H.; Doss, A.; Dhanabalan, R.; Venkataswamy, R. *Hygeia. J. D. Med.*, **2011**, *3*, 71-75.
9. ApurbaSarker Apu; Mahmuda Sulthana Liza; Tamaluddin, A.T.M.; Amran Howlader, M.D.; Repon Kumar Saha; Farhana Rizwan; Nishat Nasrin. *Asian Pacific Journal of Tropical Biomedicine.*, **2012**, *2*, 673-678.
10. <http://www.rcsb.org/pdb>
11. Schrodingersuit; **2009** Protein Preparation Wizard; Epik version 2.0; Impact version 5.5. Schrodinger, LLC, New York., **2010**.
12. Friesner, R.A.; Banks, J.L.; Murphy, R.B.; Halgren, T.A.; Klicic, J.J.; Maniz, D.T.; Repasky, M.P.; Knoll, E.H.; Shelley, M.; Perry, J.K.; Shaw, D.E.; Francis, P.; Shenkin, P.S. *J.Med.Chem.*, **2004**, *47*, 1739-1749.
13. Babitha Agarwal; Sunanda Das; Archana Pandey. *Asian journal of Applied Sciences.*, **2011**, *3*, 436-441.
14. Friesner, R.A.; Murphy, R.B.; Repasky, M.P.; Frye, L.L.; Greenwood, J.R.; Halgren, T.A.; Sanschagrín, P.C.; Maniz, D.T. *J.Med.Chem.*, **2006**, *49*, 6177-6196.
15. Schrodinger suit , Protein Preparation Guide, Site Map 2.4; Glide Version 5.6,Lig Prep 2.4,Qik Propn3.3, Schrodinger, LLC, New York, NY., **2010**.
16. Chude, M.A.; Orisakwe, O.E.; Afonne, O.J., Gamaniel, K.S.; Vongtau, O.H.; Obi, E. *Indian Journal of Pharmacology.*, **2001**, *33*, 215-216.
17. Ujowundu, C.O.; Igwe, C.U.; Enemor, V.H.A.; Nwaogu, L.A.; Okafor, O.E. *Pakistan journal of Nutrition.*, **2008**, *7*, 90-92.
18. Meera Sumanth; Mustafa, S.S. *International Journal of Pharmacology.*, **2007**, *3*, 416-420.
19. Krishnamurthy; Mayank, A.P.; Vijay Lambole. *Indian Journal of experimental biology.*, **2010**, *5*, 107-110.
20. Sharma, M.; Vohra, S.; Arnason, J.T.; Hudson, J.B. *Pharm Biol.*, **2008**, *46*, 11-116.
21. Akinnibosun, F.I.; Akinnibosun, H.A.; Ogedegbe, D. *Science World Journal.*, **2009**, *4*, 25-28.